

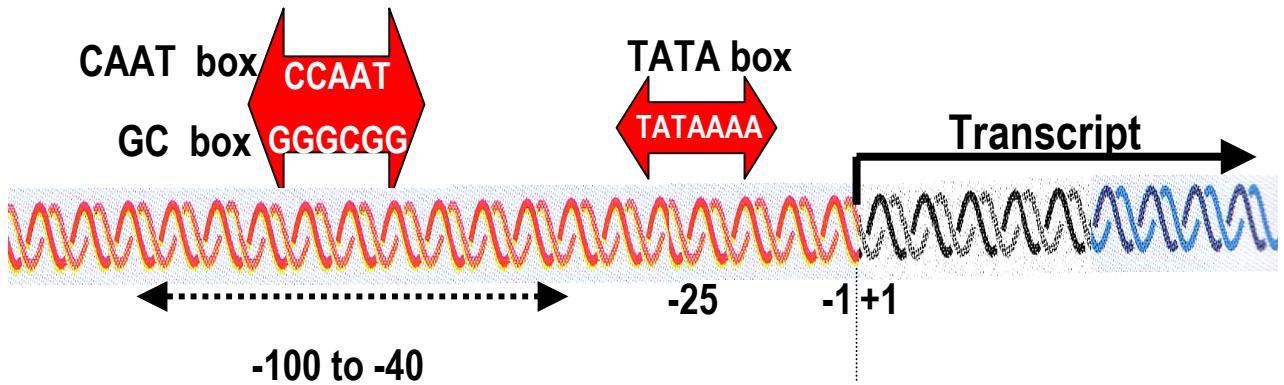
# **BIOC4004 - Industrial Biochemistry**

## **Lecture 02 - Fri Jan 09, 04**

### **Topics for the Day:**

- **Promoters**
  - **Prokaryotic vs Eukaryotic**
- **Transcription**
  - **Prokaryotic vs Eukaryotic**
- **Transcriptional regulation**
  - **Transcription factors**
- **Translation**
  - **Prokaryotic vs Eukaryotic**
- **The Genetic Code**
- **Regulation of translation**

# Generalized view of Eukaryotic Promoters

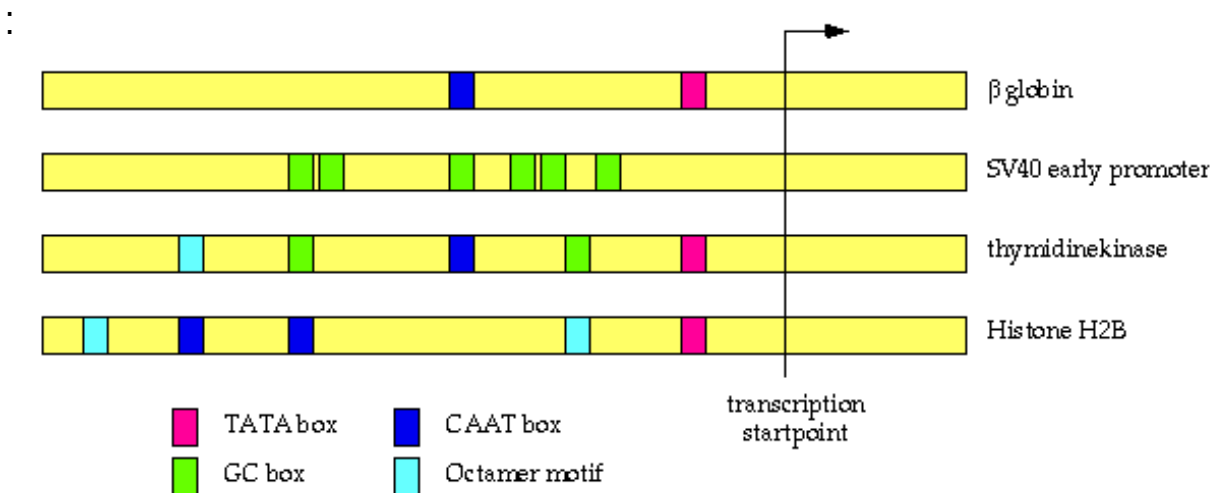


Gannon et al. (1978) looked at transcription start sites...

Chicken ovalbumin	GAGGCTATATATTC <sup>63</sup> CCCAAGGGCTCAGCCAGTGTCTGTACA
Adenovirus late	GGGGCTATAA <sup>85</sup> AGGGGGTGGGGGCGGTTTCGTCTCACTC
Rabbit β globin	TTGGGCATAA <sup>85</sup> AGGCAGAGCAGGGCAGCTGCTGCTAACACT
Mouse β globin major	GAGCATATA <sup>85</sup> AGGTGAGGTTAGGATCAGTTGCTCCTCACATT

T<sup>82</sup>A<sup>97</sup>T<sup>93</sup>A<sup>85</sup>T<sup>37</sup>
A<sup>63</sup>A<sup>83</sup>
A<sup>50</sup>T<sup>37</sup>

**note:** Some genes, including “housekeeping genes” do not have TATA boxes



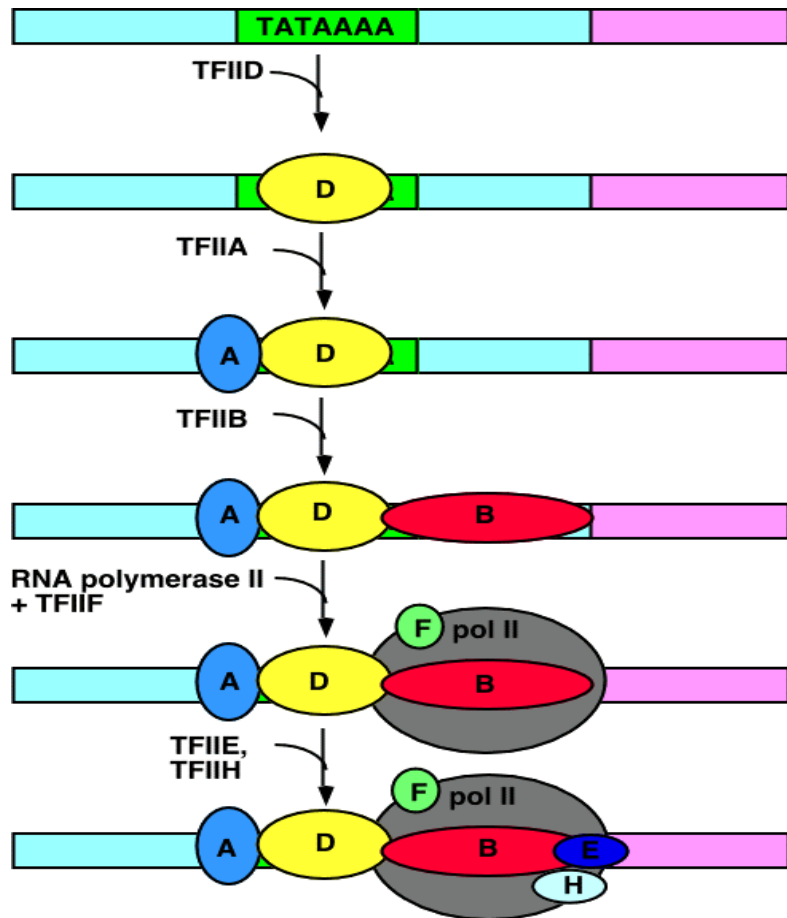
• Promoters contain combinations of “upstream regulatory motifs”

## Generalized view of Eukaryotic Promoters (cont)

- Eukaryotes have three types of RNA polymerases
- Protein-encoding genes are transcribed by **RNA pol II**
- Eukaryotic RNA polymerases are assembled from numerous subunits
  - as many as 14 different subunits come together
- At least six **general (or basal) transcription factors** have been characterized
  - TFIIA, TFIIB, TFIID, TFIIE, TFIIF, TFIIH
  - Allow “basal” transcription initiation
- At least 8 - 10 TAF<sub>II</sub>'s (TBP-associated proteins) associated with TFIID
  - the mixture of TBP-associated proteins is “highly complex”
  - Polymerases assembled at different genes differ in composition!!!

# Generalized view of Eukaryotic Promoters (cont)

## Eukaryotic Transcription



TFIID: TBP (TATA box binding protein)

TAFs (interact with regulatory sequences and their TFs)

TFIIA: stabilizes TFIID interaction with promoter

TFIIB: interaction between TFIID and PolII-TFIIF

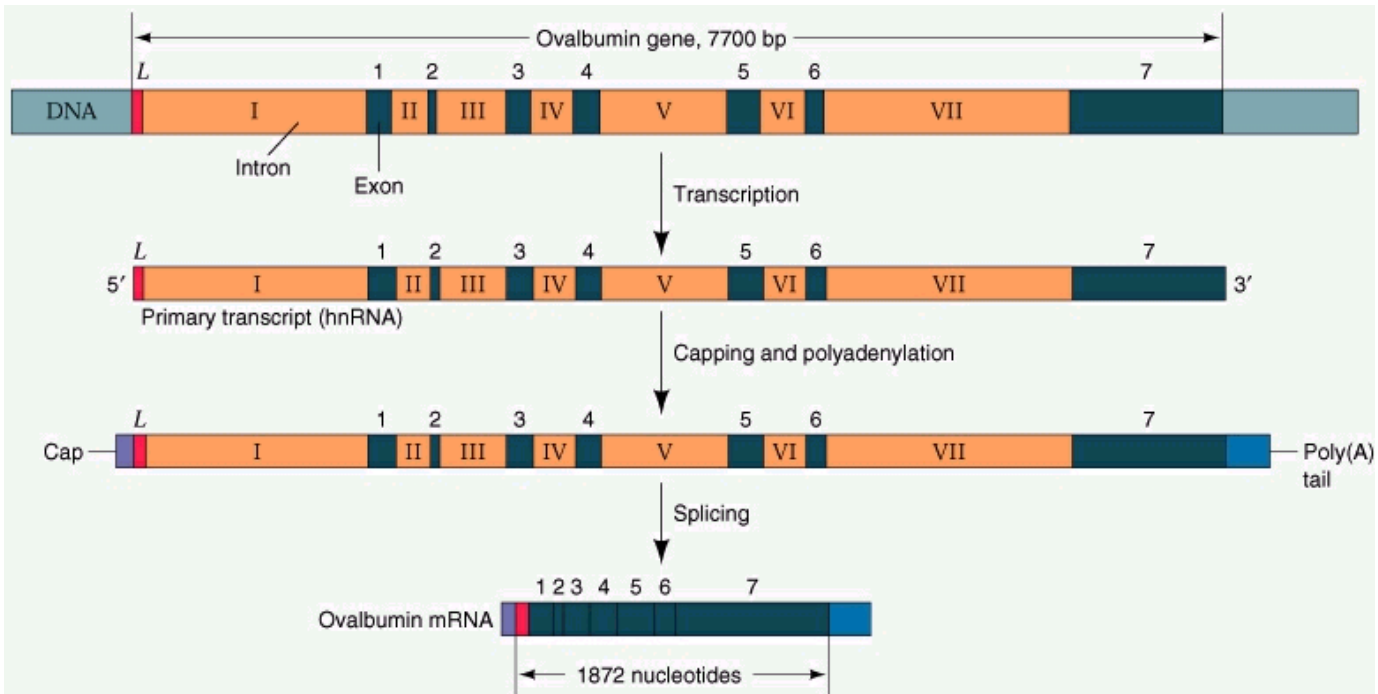
TFIIF: recruits PolII to promoter; helicase activity

TFIIE: binding and stimulation of transcription by recruiting TFIIH

TFIIH: kinase activity (activates PolII by phosphorylation - promoter clearance)

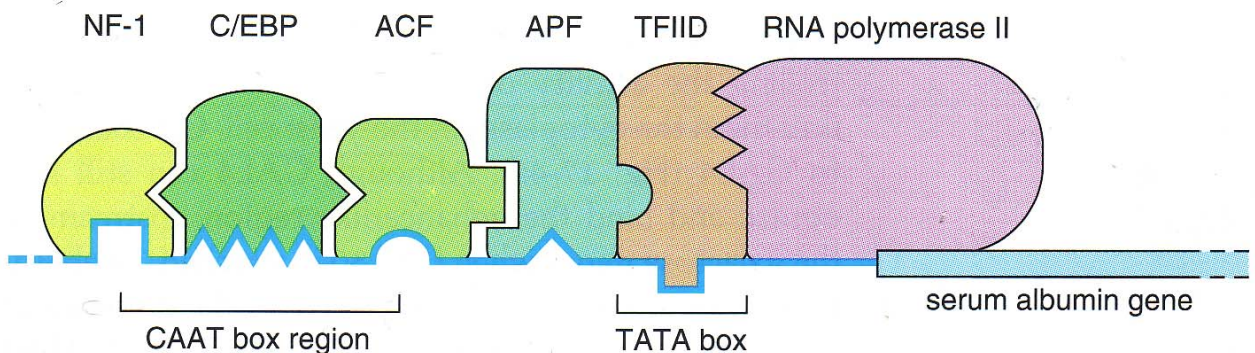
# Eukaryotic Transcription Termination:

- process still largely unknown
  - polymerase extends ~1000 bp past the mature end of the mRNA
  - termination signals cause the polymerase to fall off the DNA template
    - sequences required for termination are unknown
    - polymerase can fall off at one of several different places
- ie. a heterogeneous mixture of mRNAs of different length is produced
- mature 3'-end is generated by post-transcriptional cleavage
    - followed by polyadenylation(except for non-poly-A mRNAs)



## Generalized view of Eukaryotic Promoters (cont)

- Promoters contain combinations of “upstream regulatory motifs” to regulate expression
  - binding sites for **Transcription Factors (TFs)**
  - cooperative interaction between TFs
  - combinations allow fine tuning of gene regulation
    - temporal; tissue specific; stimulus-specific (hormone, growth factor, metabolite...)

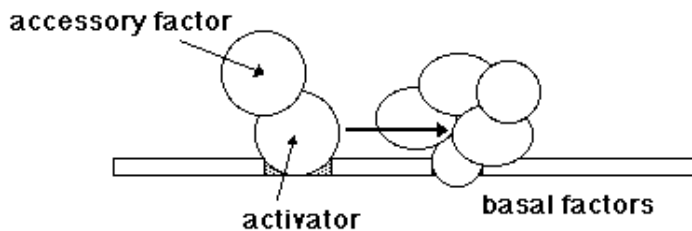
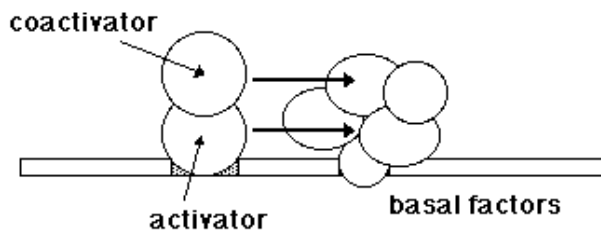
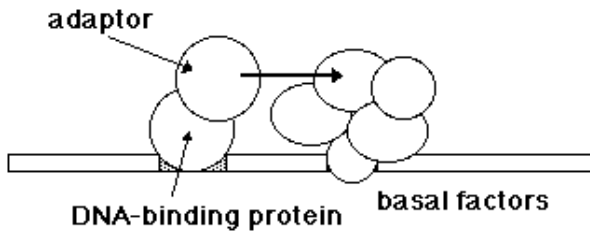


example: the Serum Albumin gene

- failure of NF-1, C/EBP or ACF to bind significantly reduces transcription
  - APF is very specific for the serum albumin gene
  - The combination of TFs increases binding affinity of RNA pol II for promoter
- 
- **Gene regulation networks** can be achieved when different genes share binding sites for a given TF

## Generalized view of Eukaryotic Promoters (cont)

Transcription Factors can come together in cooperative fashion:



Transcription Factor Activity is tightly regulated:

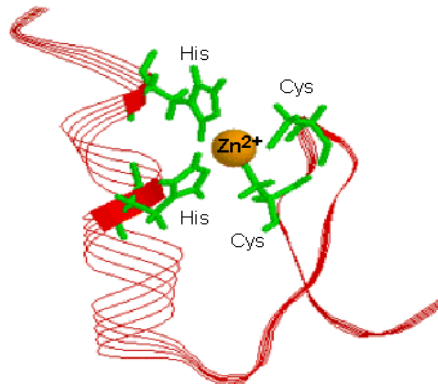
- control of TF levels
- post-translational modification (phosphorylation, dephosphorylation)
- localization
- binding to an inhibitor or activator (a molecule, a protein)
- change in oligomerization state

# A quick word on Transcription Factors

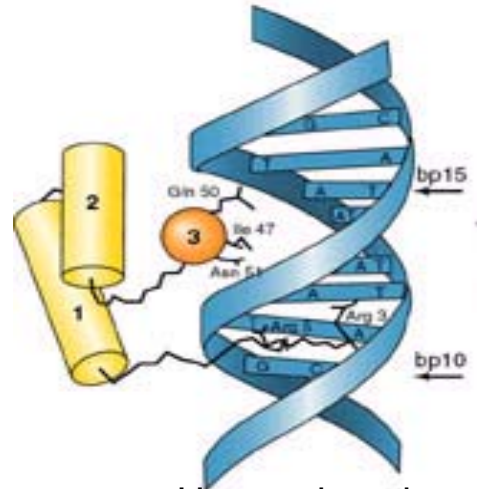
Need two things:

- DNA binding motif
- Activation (or repressing) domain

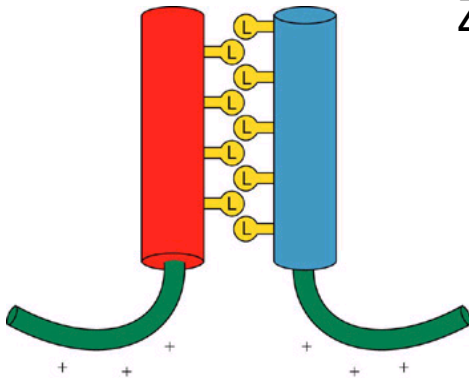
## Some DNA Binding Motifs:



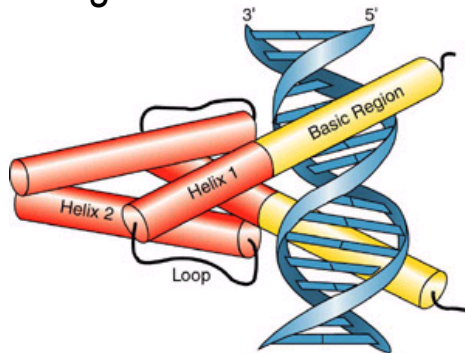
Zinc Finger



Homeodomain



Leucine Zipper



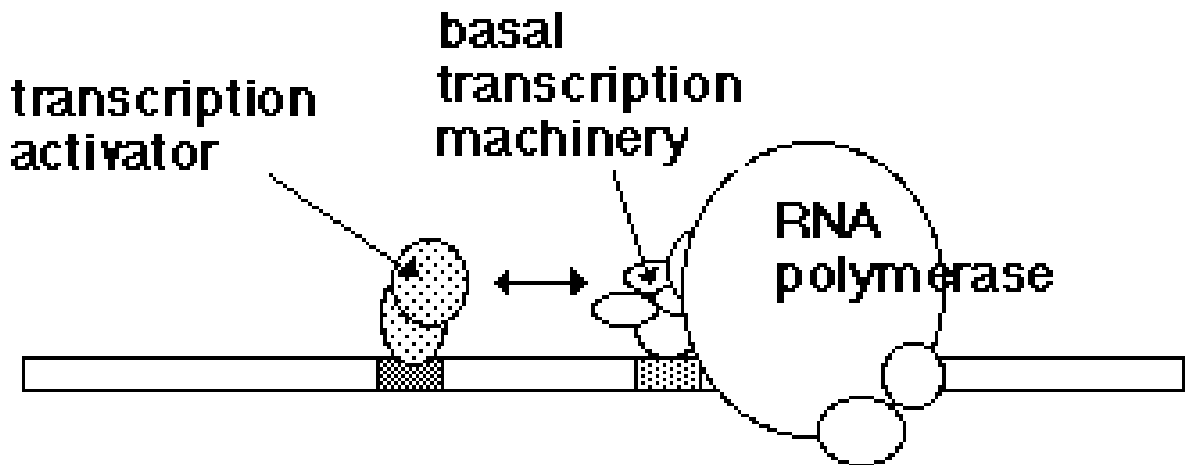
bHLH (basic Helix-Loop-Helix)

## Transcription-activating Domains

Three kinds of protein domains are observed

1. acidic domains, where the amino acid side chains are acidic in nature (glutamic acid, aspartic acid)
2. glutamine-rich domains, with about 25% glutamine in the sequence
3. proline-rich domains

## A quick word on Transcription Factors (cont)



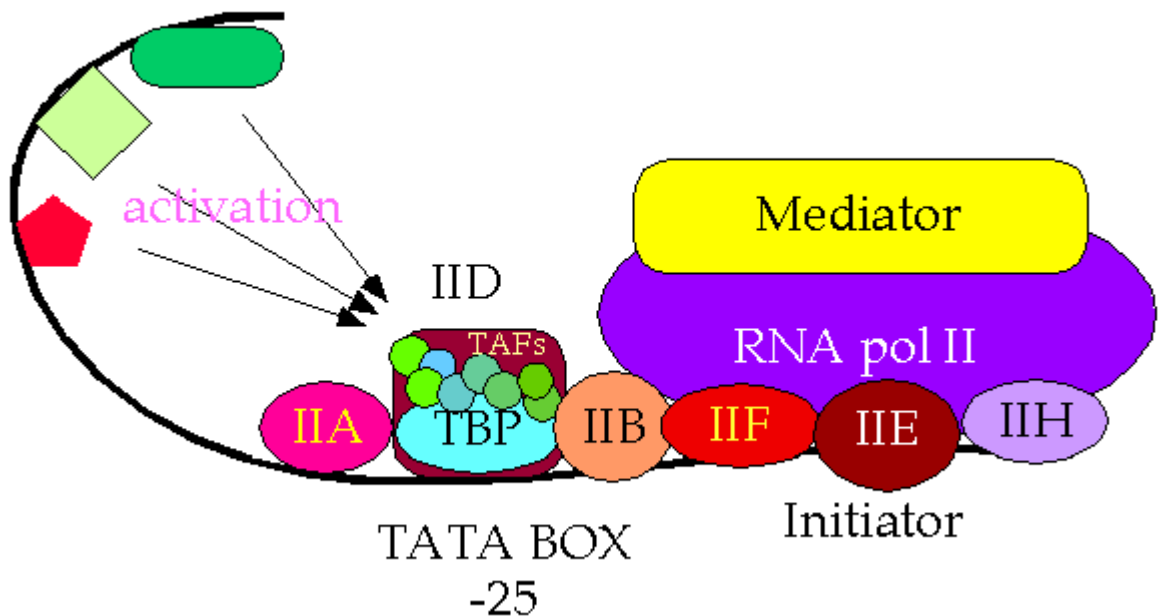
### What do Transcription Factors Do ?

- basal transcription machinery recruits RNA polymerase to the promoter
  - low to nil transcription levels
- transcription factors interact with basal factors to increase the rate of transcription
  - stimulate assembly of preinitiation complexes
  - stimulate formation of an open complex
  - stimulate promoter escape
- binding sites for transcription activators must be made accessible
  - some transcription factors function to re-model chromatin
  - remodelling makes the DNA accesible to other factors
  - “euchromatin” vs “heterochromatin”
    - DNA packaging “silences” gene expression
    - histone modification (acetylation, methylation & phosphorylation) activates DNA

# The ever-mysterious “Enhancers”

- In eukaryotic cells
  - sequences that influence the rate of transcription
  - interact with specific proteins.
  - can be located within, near, or **very far** (ie. Kbs) from the gene’s promoter
  - can be upstream or downstream of gene

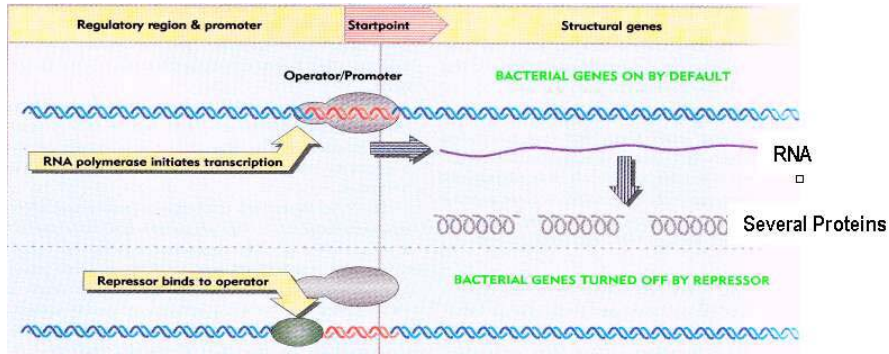
Enhancer



- Bind
- Interact with regulatory protein attached to promoter
  - DNA loop formed
- Enhance RNA pol binding

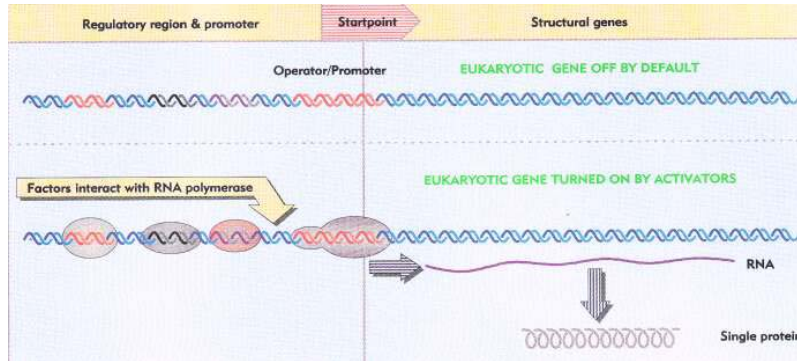
# Differences between prokaryotic and eukaryotic genes

## • Prokaryotes:



- more often than not, gene is ON by default
- most gene regulation is negative (ie. repressors proteins turn off transcription)
- a lot of related genes are arranged in operons
- coding sequences are co-linear with the encoded polypeptide (ie. no introns)
- no nucleus; mRNA can be translated as it's being transcribed

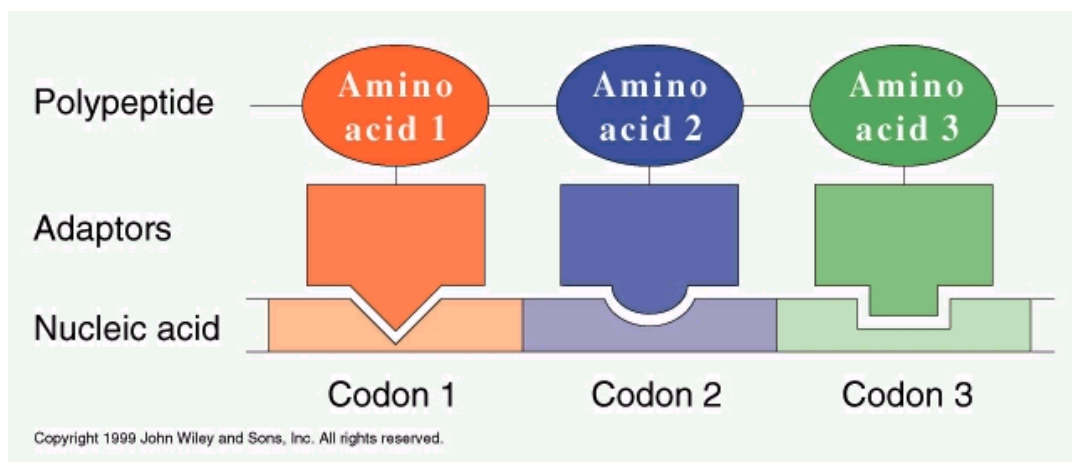
## • Eukaryotes:



- most genes are OFF by default
- most gene regulation is positive (ie. transcription factors needed for activation)
- promoters are rich with TF-binding motifs
- the majority of genes contain introns
- modifications are made before mRNAs are exported out of the nucleus
  - 5' cap
  - 3' poly[A] tail
  - splicing to remove introns (intervening sequences)
  - **alternative splicing can generate multiple polypeptides from same gene**
- translation occurs outside the nucleus:
  - secreted proteins - rough ER
  - cytosolic proteins - cytoplasm

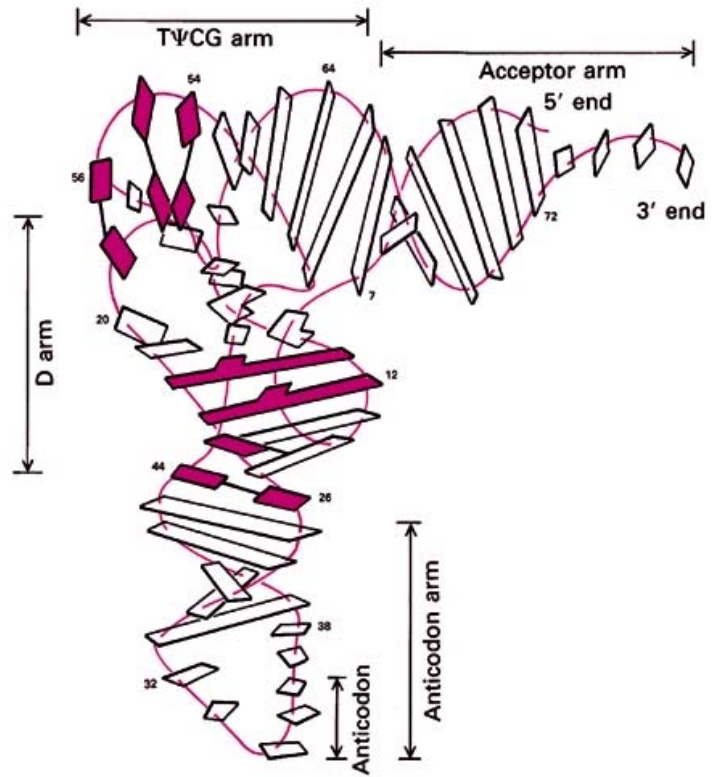
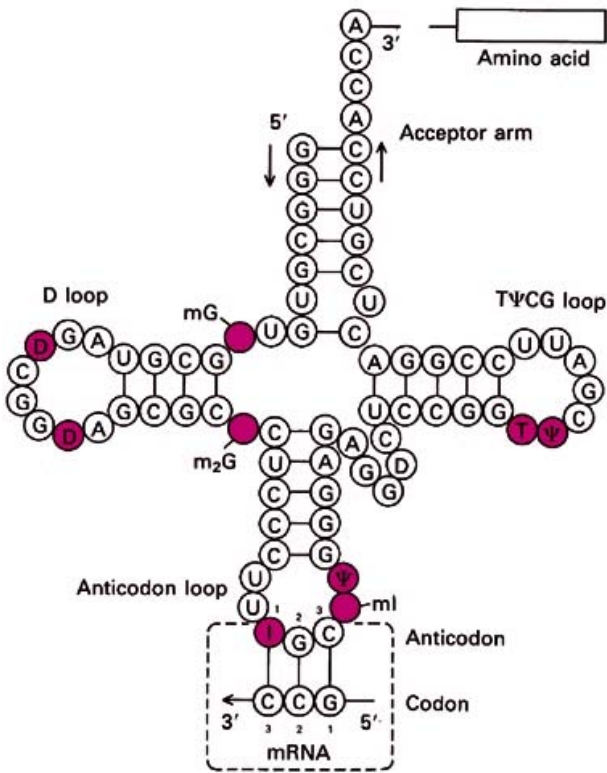
# From RNA to Protein: the process of Translation

- **Translation** decodes the mRNA sequence into the polypeptide that it encodes
- **Machinery Required**
  - mRNA
  - amino acids, tRNAs & aminoacyl tRNA synthetases
  - ribosomes & other protein factors
  - an initiator tRNA
  - Energy !!! (GTP & ATP)
- **Linking the DNA to protein synthesis**
  - FrancisCrick's "Adaptor Hypothesis"

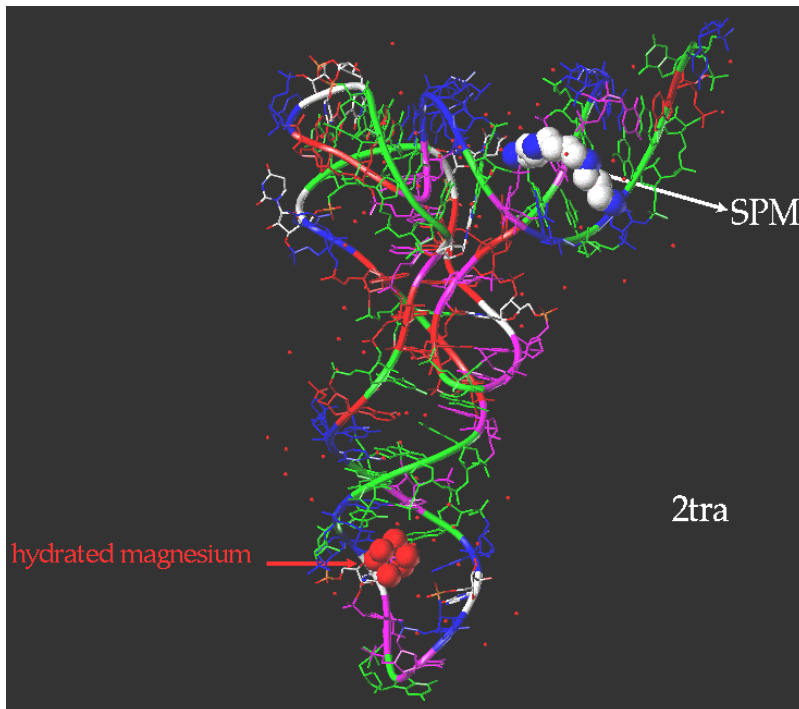


- The "adaptor" is tRNA!!!
- Specific tRNA synthetases "charge" the correct amino acid onto each specific tRNA

# A tRNA looks like this...



“An L-shaped 3-D structure”

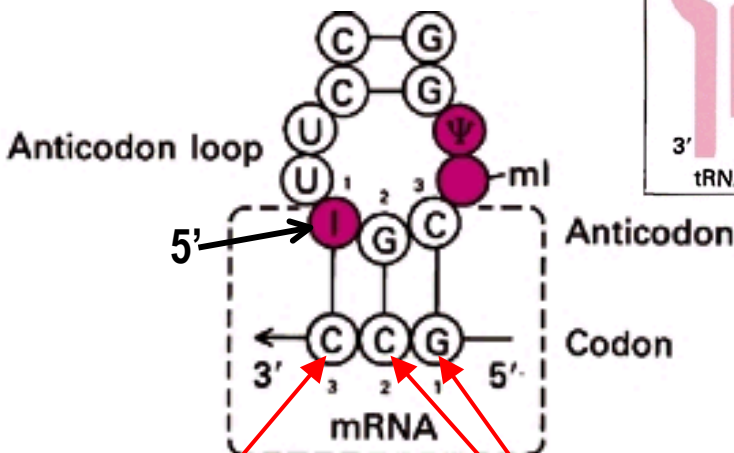
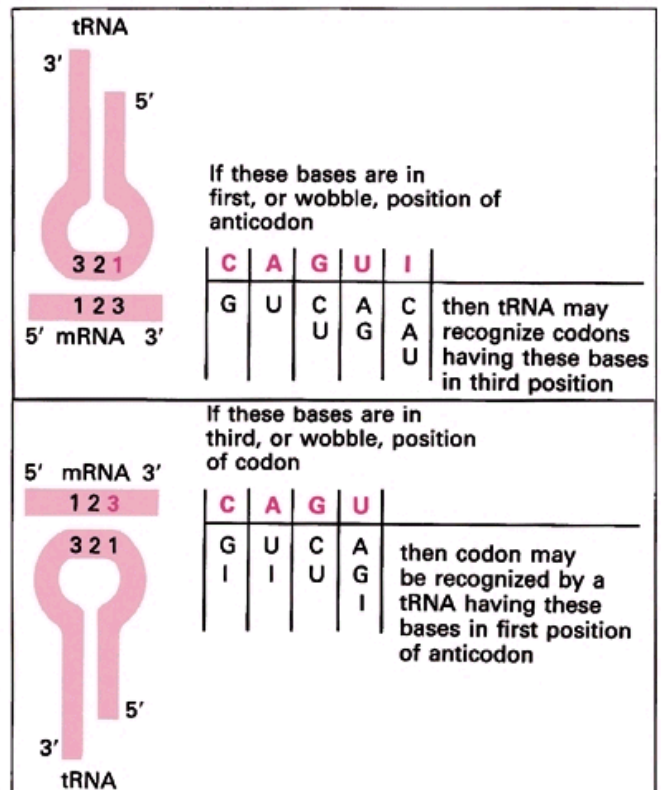


# Francis Crick's "Wobble Hypothesis"

We use 20 amino acids but have 61 codons to code for them...  
 what's up with that ????

- **Inosine** was found at the 1st (5') anticodon position (Holley, 1965)
- proposed that codon/anticodon specificity only requires 1st and 2nd codon base-pairing
- 3rd base follows non-standard base pairing with 1st anticodon position

5' anticodon base	3' codon base
A	U
C	G
G	C or U
U	A or G
I	A or C or U



- Wobble Position
- 3rd codon position
- Specificity dictated here
- 1st & 2nd codon positions

# The Genetic Code and some variants...

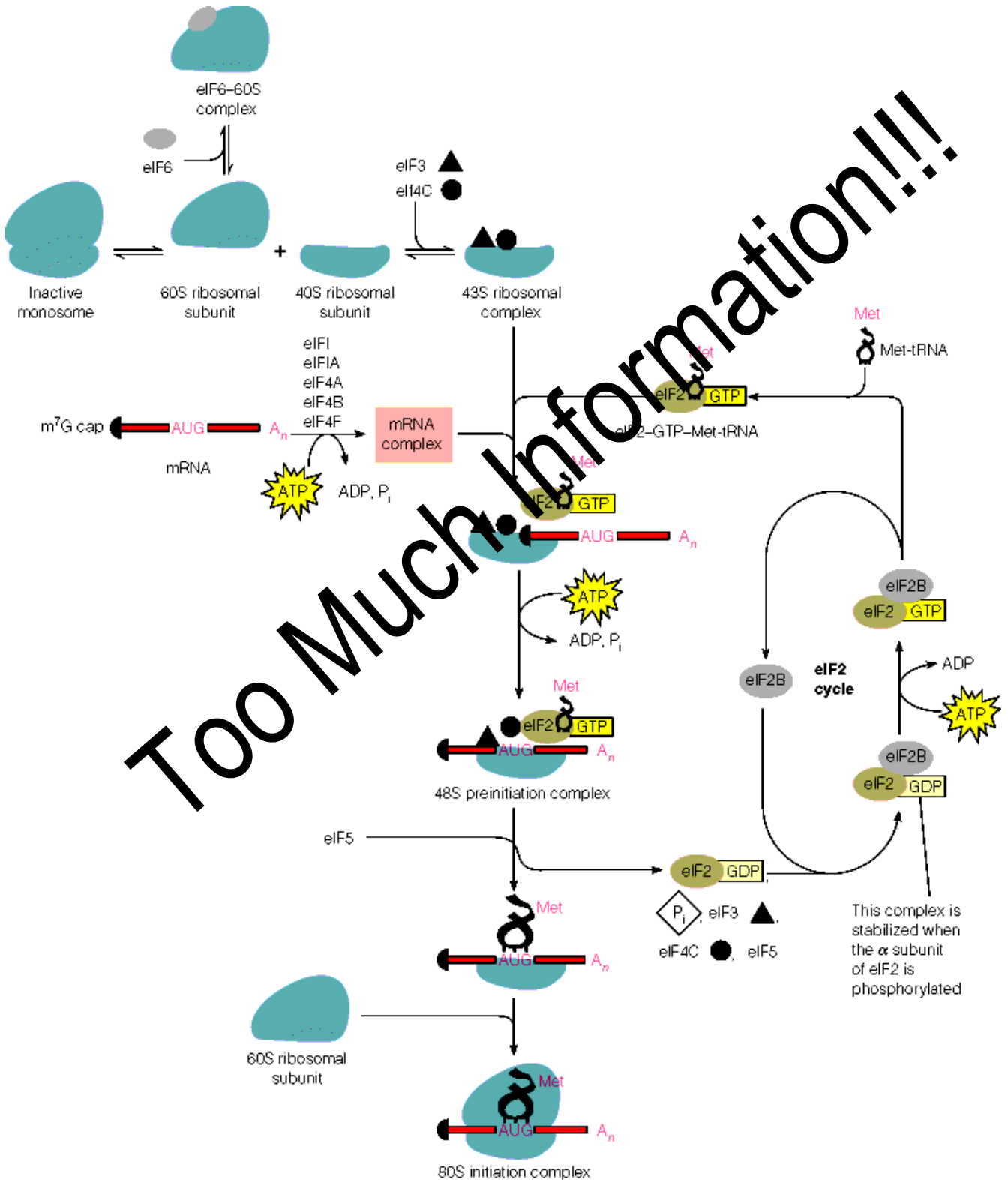
					<b>Gln</b>		<i>Diplomonads Acetabularia Some ciliates</i>
	UUU Phe UUC Phe	UCU Ser UCC Ser	UAU Tyr UAC Tyr	UGU Cys UGC Cys			
	UUA Leu UUG Leu	UCA Ser UCG Ser	<b>UAA TER</b> <b>UAG TER</b>	<b>UGA TER</b> UGG Trp			<b>Cys / Trp</b> <i>Euploides/ Mycoplasma Spiroplasma</i>
	CUU Leu CUC Leu	CCU Pro CCC Pro	CAU His CAC His	CGU Arg CGC Arg			
<b>Ser</b> <i>Candida - Saccharomyces</i>	CUA Leu CUG Leu	CCA Pro CCG Pro	CAA Gln CAG Gln	CGA Arg <b>CGG Arg</b>			<b>Nonsense</b> <i>Mycoplasma Spiroplasma</i>
	AUU Ile AUC Ile	ACU Thr ACC Thr	AAU Asn AAC Asn	AGU Ser AGC Ser			
<b>Nonsense</b> <i>Microbococcus</i>	AUA Ile <b>AUG Met</b>	ACA Thr ACG Thr	AAA Lys AAG Lys	<b>AGA Arg</b> <b>AGG Arg</b>			<b>Nonsense</b> <i>Microbococcus</i>
	GUU Val GUC Val	GCU Ala GCC Ala	GAU Asp GAC Asp	GGU Gly GGC Gly			
	GUA Val GUG Val	GCA Ala GCG Ala	GAA Glu GAG Glu	GGA Gly GGG Gly			

## The Genetic Code:

- Based on triplets(codons)
- Codons:
  - no gaps between them
  - are non-overlapping
  - unambiguous (ie. a codon codes for a single amino acid)
- **The code is degenerate:**
  - most amino acids are specified by more than one codon
  - all of the degeneracy is found at the 3rd codon position (Wobble position)
- The code is “universal” (though there are exceptions)

Great implications for genetic engineering !!! Why ?

# Eukaryotic Translation:



Too Much Information!!!

# Translation, the important factoids....

## The Mechanics are similar for Prokaryotes and Eukaryotes:

- mRNAs are read from 5' to 3'
- Proteins are synthesized from Amino- to Carboxy- terminus
  
- **Machinery Required**
  - mRNA
  - amino acids, tRNAs & aminoacyl tRNA synthetases
  - ribosomes & other protein factors
  - an initiator tRNA
  - Energy !!! (GTP & ATP)
  
- **Initiation**
  - detection of translation start site
  - assembly of the ribosome on the mRNA
  - initiator tRNA-met is brought in
  
- **Elongation**
  - charged aminoacyl-tRNA brought in
  - new peptide bond gets made
  - ribosome translocates down three positions along mRNA
  - cycle repeats until stop codon is reached
  
- **Termination**
  - binding of release factors
  - release of the completed polypeptide
  - disassembly of the ribosome from the mRNA

# Regulation of Translation

- Ribosome binding is a rate-limiting step
- Ability to initiate translation
- mRNA stability (short in Prokaryotes vs long in Eukaryotes)
- Secondary structure in the 5' UTR can inhibit translation
- Rate of translation
- Rate of re-initiation
- Presence of binding sites for proteins that can block translation
  - Iron Response Element (IRE) & IRE-BP
- Use of “Rare Codons”
  - most organisms do not encode every possible tRNA gene in their genomes
    - certain codons are not used at all !!!! Either made in very low amounts, or not at all

## • Codon Bias

- abundance of specific tRNAs (or of “charged” tRNAs) can influence translation efficiency

## • Codon Adaptive Index

- implication for heterologous gene expression

FBA1	ENO2	PYK1	FBA1	ENO2	PYK1		
TTT	0	1	0	TCT	11	13	13
TTC	15	14	15	TCC	8	17	13
TTA	7	2	3	TCA	0	0	1
TTG	17	36	32	TCG	0	0	0
CTT	0	0	0	CCT	0	0	1
CTC	0	0	0	CCC	0	0	0
CTA	0	0	0	CCA	16	13	24
CTG	0	0	0	CCG	0	0	0
ATT	8	11	11	ACT	10	8	10
ATC	14	9	26	ACC	10	12	28
ATA	1	0	0	ACA	0	0	0
ATG	8	10	11	ACG	0	0	0

Proline codon bias in three different *C. albicans* genes

# Protein Synthesis: Prokaryotes vs. Eukaryotes

- **Main differences:**

- Eukaryotic ribosomes are larger
- Prokaryotic mRNAs have significantly shorter half-lives (min vs hrs)
- Eukaryotes require **many** more factors for initiation

- **Prokaryotes:**

- ribosomal binding results from 18S subunit binding to the Shine-Delgarno motif (Ribosomal binding site)
- First MET is formylated
- no mRNA modifications
- no nucleus; translation is **concurrent** with transcription
- polycistronic

- **Eukaryotes:**

- Ribosomal binding in Eukaryotes results from 5' Cap recognition
  - In a few cases, IRES (internal ribosome entry sites)
- ribosomal scanning to correct ATG
  - Kozak motif - (GCC)(GCC)GCCGCC**ATGG**
- need splicing, 5' capping, poly-adenylation
- done inside the nucleus (concurrent with transcription)
- transcription in nucleus, translation outside nucleus
  - export mature mRNA before translation can happen
- monocistronic (however, alternative splicing & polyadenylation)

Next Time:

- protein folding
- post-translational modification
- Targetting and degradation
- quick overview of basic protein structure
  
- Recombinant DNA methodologies